RESEARCH ARTICLE

The genetic and epigenetic profile of serum S100 β in the Lothian Birth Cohort 1936 and its relationship to Alzheimer's disease [version 1; peer review: 2 approved]

Danni A Gadd ^{1*}, Robert I McGeachan ^{2*}, Robert F Hillary ^{1*}, Daniel L McCartney¹, Sarah E Harris ^{3,4}, Roy A Sherwood⁵, N Joan Abbott⁶, Simon R Cox ^{3,4}, Riccardo E Marioni¹

¹Institute of Genetics and Cancer, University of Edinburgh, Edinburgh, Other (Non-U.S.), EH4 2XU, UK

²Centre for Discovery Brain Sciences, University of Edinburgh, Edinburgh, Other (Non-U.S.), EH8 9JZ, UK

³Lothian Birth Cohorts, University of Edinburgh, Edinburgh, Other (Non-U.S.), EH8 9JZ, UK

⁴Department of Psychology, University of Edinburgh, Edinburgh, Other (Non-U.S.), EH8 9JZ, UK

⁵Department of Clinical Biochemistry, King's College Hospital NHS Foundation Trust, London, Other (Non-U.S.), SE5 9RS, UK

⁶Institute of Pharmaceutical Science, King's College London, London, Other (Non-U.S.), WC2R 2LS, UK

* Equal contributors

V1 First published: 10 Nov 2021, 6:306
<https://doi.org/10.12688/wellcomeopenres.17322.1>

Latest published: 27 Jan 2022, 6:306
<https://doi.org/10.12688/wellcomeopenres.17322.2>

Abstract

Background: Circulating S100 calcium-binding protein (S100 β) is a marker of brain inflammation that has been associated with a range of neurological conditions. To provide insight into the molecular regulation of S100 β and its potential causal associations with Alzheimer's disease, we carried out genome- and epigenome-wide association studies (GWAS/EWAS) of serum S100 β levels in older adults and performed Mendelian randomisation with Alzheimer's disease.

Methods: GWAS (N=769, mean age 72.5 years, sd = 0.7) and EWAS (N=722, mean age 72.5 years, sd = 0.7) of S100 β levels were performed in participants from the Lothian Birth Cohort 1936. Conditional and joint analysis (COJO) was used to identify independent loci. Expression quantitative trait locus (eQTL) analyses were performed for lead loci that had genome-wide significant associations with S100 β . Bidirectional, two-sample Mendelian randomisation was used to test for causal associations between S100 β and Alzheimer's disease. Colocalisation between S100 β and Alzheimer's disease GWAS loci was also examined.

Results: We identified 154 SNPs from chromosome 21 that associated ($P < 5 \times 10^{-8}$) with S100 β protein levels. The lead variant was located in the *S100 β* gene (rs8128872, $P = 5.0 \times 10^{-17}$). We found evidence that two independent causal variants existed for both transcription of *S100 β* and S100 β protein levels in our eQTL analyses. No CpG sites were

Open Peer Review

Approval Status

	1	2
version 2 (revision) 27 Jan 2022		
version 1 10 Nov 2021	 view	 view

1. **Nicholas W. Wood**, University College London, London, UK

2. **Manu Sharma** , University of Tübingen, Tübingen, Germany

Any reports and responses or comments on the article can be found at the end of the article.

associated with $S100\beta$ levels at the epigenome-wide significant level ($P < 3.6 \times 10^{-8}$); the lead probe was cg06833709 ($P = 5.8 \times 10^{-6}$), which mapped to the *LG11* gene. There was no evidence of a causal association between $S100\beta$ levels and Alzheimer's disease or vice versa and no evidence for colocalisation between $S100\beta$ and Alzheimer's disease loci.

Conclusions: These data provide insight into the molecular regulators of $S100\beta$ levels. This context may aid in understanding the role of $S100\beta$ in brain inflammation and neurological disease.

Keywords

Epigenetic, Genetic, $S100\beta$, Inflammation, EWAS, GWAS, Alzheimer's disease

This article is included in the [Generation Scotland gateway](#).

Corresponding authors: Danni A Gadd (danni.gadd@ed.ac.uk), Riccardo E Marioni (Riccardo.Marioni@ed.ac.uk)

Author roles: **A Gadd D:** Conceptualization, Data Curation, Formal Analysis, Investigation, Methodology, Project Administration, Software, Validation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; **I McGeachan R:** Formal Analysis, Investigation, Methodology, Visualization, Writing – Original Draft Preparation; **F Hillary R:** Formal Analysis, Investigation, Methodology, Writing – Original Draft Preparation; **L McCartney D:** Data Curation, Formal Analysis, Methodology; **E Harris S:** Data Curation, Project Administration; **A Sherwood R:** Data Curation, Resources; **Abbott NJ:** Data Curation, Resources; **R Cox S:** Data Curation, Funding Acquisition, Project Administration; **E Marioni R:** Conceptualization, Data Curation, Formal Analysis, Funding Acquisition, Investigation, Methodology, Project Administration, Resources, Software, Supervision, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: R.E.M. has received payment from Illumina for presentations and is an advisor to the Epigenetic Clock Development Foundation. All other authors declare no competing interests.

Grant information: This research was funded in whole, or in part, by Wellcome [104036/Z/14/Z, 108890/Z/15/Z, 203771/Z/16/Z]. For the purpose of open access, the author has applied a CC BY public copyright licence to any Author Accepted Manuscript version arising from this submission. The LBC1936 is supported by Age UK (Disconnected Mind project, which supports S.E.H.), the Medical Research Council (G0701120, G1001245, MR/M013111/1, MR/R024065/1), and the University of Edinburgh. Genotyping was supported by the Biotechnology and Biological Sciences Research Council (BB/F019394/1). Methylation typing in the LBC1936 was supported by Centre for Cognitive Ageing and Cognitive Epidemiology (Pilot Fund award), Age UK, The Wellcome Trust Institutional Strategic Support Fund, The University of Edinburgh, and The University of Queensland. This work was supported by the Medical Research Council and Biotechnology and Biological Sciences Research Council (MR/K026992/1). S.R.C. is also supported by a Sir Henry Dale Fellowship jointly funded by the Wellcome Trust and the Royal Society (Grant Number 221890/Z/20/Z). D.A.G. and R.F.H., are supported by funding from the Wellcome Trust 4-year PhD in Translational Neuroscience–training the next generation of basic neuroscientists to embrace clinical research [108890/Z/15/Z]. R.I.M. is supported by funding from the Wellcome Trust PhD for clinicians, Edinburgh Clinical Academic Track for Veterinary Surgeons. D.L.M.C. and R.E.M. were supported by Alzheimer's Research UK major project grant ARUK-PG2017B–10. R.E.M. is supported by Alzheimer's Society major project grant AS-PG-19b-010.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Copyright: © 2021 A Gadd D *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: A Gadd D, I McGeachan R, F Hillary R *et al.* **The genetic and epigenetic profile of serum $S100\beta$ in the Lothian Birth Cohort 1936 and its relationship to Alzheimer's disease [version 1; peer review: 2 approved]** Wellcome Open Research 2021, 6:306 <https://doi.org/10.12688/wellcomeopenres.17322.1>

First published: 10 Nov 2021, 6:306 <https://doi.org/10.12688/wellcomeopenres.17322.1>

Introduction

The calcium-binding protein S100 beta (S100 β) has been suggested as a biomarker for central nervous system disease¹. Expressed most commonly in astrocytes, its cytoplasmic location and calcium-binding capability allows S100 β to mediate calcium homeostasis, cell proliferation and survival intracellularly, while also triggering the RAGE-associated inflammatory response when secreted extracellularly². Part of pro-inflammatory danger-associated molecular patterns (DAMPs), elevated S100 β is linked to cytokine cascades in the brain^{3,4}.

Although the exact pathophysiology is still unknown, a number of small-scale studies have reported elevated circulating or cerebrospinal fluid (CSF) S100 β levels in individuals with nervous system injury, neuroinflammatory conditions, white matter ageing and Alzheimer's dementia and delirium⁴⁻⁹. Following traumatic brain injury, elevated S100 β plasma levels have been shown to precede increases in intracranial pressure¹. Blood-based levels of S100 β have also been found to be elevated in small vessel disease and associated with cognitive decline^{10,11}. Activated astrocytes expressing high levels of S100 β have been profiled at plaques in the hippocampus, temporal lobe, frontal lobe and pons in individuals with Alzheimer's disease¹². The *S100 β* gene was also identified as a site of differential DNA methylation (DNAm) relating to Braak staging in a previous epigenome-wide association study (EWAS) of cortical post-mortem tissues (n=159)¹³.

Whether S100 β has a direct involvement in the pathogenesis of Alzheimer's disease is still unclear. Further to its potential role in inflammatory exacerbation in the brain, research suggests that at nanomolar concentrations, S100 β can have protective and neurotrophic effects¹⁴. Despite the widely discussed biological importance of S100 β , the possible epigenetic regulators of the protein have not been investigated. One study using blood spots taken at birth has previously identified two genetic associations in relation to circulating S100 β levels; rs62224256 on chromosome 21, 21kb downstream of the pericentrin gene (*PCNT*) and rs28397289 on chromosome 6, within the human leukocyte antigen (*HLA*) region¹⁵. Elucidating the mechanisms that determine inter-individual variation in circulating S100 β levels, in healthy individuals, may therefore provide insight

into S100 β 's role in health and disease. Further genetic mapping of S100 β may also facilitate causal association tests with disease endpoints.

Here, we perform genome- and epigenome-wide association studies (GWAS/EWAS) of S100 β in relatively healthy older adults from the Lothian Birth Cohort 1936 (LBC1936). We then use genetic instruments identified for S100 β to test for bidirectional causal associations with Alzheimer's disease, via two-sample Mendelian randomisation. Demographic information for the GWAS (N=769) and EWAS (N=722) sample groups are summarised in Table 1.

Methods

The Lothian Birth Cohort 1936

The Lothian Birth Cohort 1936 (LBC1936) is a longitudinal study of cognitive ageing. Cohort members were born in 1936 and took part in the Scottish Mental Survey 1947 at age 11 years. Approximately 60 years later, those individuals that were living mostly within the Edinburgh area were re-contacted (n = 1,091, recruited at mean age 70 years). Recruitment and testing of the LBC1936 cohort have been described previously^{16,17}. Briefly, the data collection has included detailed phenotypic, biological and cognitive sampling, over a series of follow-up waves (approximately every 3–4 years since recruitment). Body mass index (BMI) was measured at clinic visits and recorded in kg/m². The S100 β data available in this study were from the second wave of testing, at around three years after the Wave 1 visit to the clinic. DNA methylation data were also collected at the same time-point as S100 β sampling. Genotyping was performed on DNA from blood samples collected at Wave 1.

S100 β measurement

Serum samples were obtained from participants during the main physical and cognitive testing appointment at Wave 2. After collection, samples were stored at –80 °C at the Wellcome Trust Clinical Research Facility, Western General Hospital, Edinburgh, until the conclusion of the wave. They were then transferred to the Department of Clinical Biochemistry, King's College London using cold-chain logistics, where they were stored at –20 °C until assays were conducted using a chemiluminescence immunoassay S100 β kit (catalogue number 314701,

Table 1. Demographics summary for the EWAS and GWAS sample populations.

Phenotype	EWAS		GWAS	
	n (%)	Mean (sd)	n (%)	Mean (sd)
Maximum sample (n)	722		769	
S100 β (μ g/L)		0.086 (0.035)		0.085 (0.034)
Sex (Female)	340 (47.1)		367 (47.7)	
Age at S100 β (years)		72.5 (0.7)		72.5 (0.7)
Body Mass Index (kg/m ²)		27.9 (4.3)		27.9 (4.3)
EpiSmokEr		0.84 (5.2)		

distributed by DiaSorin, Berks, UK) on a LIAISON chemiluminescence analyzer. The lag between sample dispatch at the end of sampling and assay completion (i.e., time stored at -20°C rather than -80°C) was an average of 44 days (SD = 26) for four batches. The minimal detectable concentration of the assay was $0.02\text{ }\mu\text{g/L}$.

Genotyping

LBC1936 DNA samples were genotyped at the Edinburgh Clinical Research Facility using the Illumina 610-QuadV1 array (Wave 1; $n = 1005$; mean age: 69.6 ± 0.8 years; San Diego). Preparation and quality control steps have been reported previously¹⁸. SNPs were imputed to the 1000 G reference panel (phase 1, version 3). Briefly, individuals were excluded on the basis of sex mismatches, relatedness, SNP call rate of less than 0.95, and evidence of non-European ancestry. SNPs with a call rate of greater than 0.98, minor allele frequency in excess of 0.01, and Hardy-Weinberg equilibrium test with $P \geq 0.001$ were included in analyses. Only SNPs with a minor allele frequency > 0.05 and imputation quality > 0.6 were retained. The remaining 8,489,963 SNPs were filtered to remove those that had a minor allele count < 25 . A total of 7,307,523 SNPs were available for GWAS analyses.

DNA methylation

DNA methylation from whole blood at Wave 2 of the Lothian Birth Cohort 1936 was measured using the Illumina 450 K methylation array at the Edinburgh Clinical Research Facility. Complete details of quality control steps taken to process the dataset have previously been described¹⁹. Briefly, raw intensity data were background-corrected and normalised using internal controls. Manual inspection facilitated the removal of low quality samples presenting issues relating to bisulphite conversion, staining signal, inadequate hybridisation or nucleotide extension. Further quality control analyses were performed to exclude probes with low detection rate $< 95\%$ at $P < 0.01$ and samples with a low call rate ($< 450,000$ probes detected at p -values of less than 0.01) were also excluded. Finally, samples were removed if there was a poor match between genotype and incorrect DNA methylation-predicted sex, or SNP control probes. DNA methylation at Wave 2 was processed in three sets ($n=256$, 461 and 5, for sets 1, 2, and 3, respectively). In total, there were 459,309 CpG sites used in EWAS analyses. Lothian Birth Cohort Wave 2 DNA methylation data were used to generate an epigenetic score for smoking – known as EpiSmokEr²⁰ in the sample. This score utilises previously derived weights calculated in an independent sample and has been previously shown to robustly reflect smoking status²⁰.

S100 β sample preparation

Of the 834 individuals with S100 β concentrations recorded at Wave 2, there were 769 individuals with genome-wide genetic data (mean 72.5 years, $sd = 0.7$) and 722 individuals with epigenome-wide DNA methylation data (mean 72.5 years, $sd = 0.7$) available. Table 1 summarises demographic information for these sample populations. In the maximum sample available in GWAS and EWAS ($N=722$), serum S100 β levels

were higher in females ($\beta = 0.26$, $SE = 0.07$, $P = 2.2 \times 10^{-4}$) and older individuals (correlation of 0.18 and $\beta = 0.16$ per year, $SE = 0.03$, $P = 2.9 \times 10^{-6}$ in linear models) (Figure 1).

Body mass index (BMI) and smoking are common lifestyle covariates that have well-documented DNAm signatures^{21,22}; we therefore tested whether these traits should be adjusted for in our analyses. S100 β levels were positively associated with body mass index (BMI) ($\beta = 0.10$ per kg/m^2 , $SE = 0.03$, $P = 2 \times 10^{-3}$), but did not associate with the DNAm-based score for smoking, EpiSmokEr ($\beta = -0.01$ per unit increase in the DNAm smoking score, $SE = 0.04$, $P = 0.78$). For this reason, BMI was included as a covariate in all analyses.

S100 β protein levels were transformed by rank-based inverse normalisation and regressed onto age, sex, BMI at Wave 2 (kg/m^2) and four genetic principal components of ancestry in separate analyses groups (EWAS $N=722$, GWAS $N=769$). Standardised residuals (mean = 0, variance = 1) from these linear regression models were brought forward as the protein level variable for the respective analyses.

Data quality control and preparation was conducted in R (Version 4.0.3)²³

Genome-wide association study (GWAS)

Linear regression was used to assess the effect of each of the 7,307,523 available SNPs on the levels of S100 β via PLINK (Version 1.9)²⁴. Genome-wide stepwise conditional analysis was performed through GCTA-COJO using the ‘cojo-slc’ option to identify independent variants. Individual level genotype data were used for the reference linkage disequilibrium (LD) structure along with default settings of the software²⁵.

Expression quantitative trait loci (eQTL) colocalisation

We cross-referenced sentinel *cis* pQTLs that were selected by GCTA-COJO analyses with publicly available *cis* eQTL data taken from the eQTLGen consortium²⁶. The *cis* eQTLs were subset to the same chromosome as the *cis* pQTL. A 200 kb region (either upstream or downstream) was extracted from our GWAS summary statistics for S100 β to capture *cis* effects within 100 kb of the target gene²⁷. eQTLs for this region were then extracted from the eQTLGen consortium summary statistics for the S100 β region. The shared SNPs across transcripts for S100 β and S100 β protein levels were then tested for colocalisation using the coloc package²⁸ (Version 5.1.0) in R, with five hypotheses in Bayesian tests with default priors²⁸. In addition to the null hypothesis (no causal variant), hypothesis 1 indicated a causal variant was present for S100 β protein levels only. Hypothesis 2 indicated that there was a causal variant for the S100 β transcript only. Hypothesis 3 indicated that there were independent causal variants for both S100 β transcription and S100 β protein levels. Hypothesis 4 indicated that there were two association signals that contributed to both S100 β gene expression and S100 β protein levels. A posterior inclusion probability (PP) > 0.95 was taken as the threshold for hypothesis testing.

Figure 1. S100β in Wave 2 of the Lothian Birth Cohort 1936 (N=722). **A**, Violin plot to illustrate differences in S100β by sex. **B**, Scatterplot with regression line and 95% CIs for S100β by age at serum sampling with a Pearson's correlation coefficient (r) annotated. S100β is plotted in μg/L units in all instances and age is given in years.

Mendelian randomisation and colocalisation

Two-sample, bidirectional Mendelian Randomisation (MR) was used to test for causal associations between S100β protein levels and neurological diseases. Summary statistics from a GWAS performed by Jansen *et al.*²⁹ were used as the Alzheimer's disease dataset (13,367,299 SNPs, with N=71,880 cases, and N=383,378 controls). All analyses were performed using the TwoSampleMR package (Version 0.5.6) in R³⁰. Clumping was used to prune SNPs for linkage disequilibrium (LD) at $r^2 < 0.001$. When testing causal associations with S100β as the exposure and Alzheimer's disease as the outcome, only one of the 154 SNPs with $P < 5 \times 10^{-8}$ (rs8128872; the sentinel variant identified by GCTA-COJO analyses in our GWAS of S100β) remained after LD pruning. The causal effect estimate for S100β to Alzheimer's disease was therefore determined using the Wald ratio test. An F statistic for the strength of the association between the sentinel SNP and the exposure was calculated using the method: $F = ((N-k-1) / k) \times (r^2 / (1-r^2))$, where N = sample size, k = number of SNPs and r^2 = variance explained in S100β levels by the genetic instruments. The r^2 statistic was calculated as follows: $r^2 = 2 \times \text{MAF} \times (1-\text{MAF}) \times \text{beta}^2$, where beta = effect size of the SNP and MAF = the effect allele frequency. When testing causal associations with Alzheimer's disease as the exposure and S100β as the outcome, 42 of the 2,357 SNPs with $P < 5 \times 10^{-8}$ remained after LD pruning and 29 were present in the S100β summary statistics. Multi-method Mendelian randomisation was then performed using the 29

SNPs from the Jansen *et al.*²⁹ summary statistics using the TwoSampleMR package in R.

The coloc package (Version 5.1.0) was used to conduct colocalisation analyses for the sentinel SNP in the S100β region and the Jansen *et al.*²⁹ summary statistics for Alzheimer's disease GWAS. Each dataset was subset to a 200 kb section (upstream or downstream) surrounding the sentinel SNP on chromosome 21. Four hypotheses were used in Bayesian tests with default priors²⁸, as per the eQTL colocalisation tests, but for the presence of Alzheimer's disease causal variants in the same region as S100β. A posterior inclusion probability (PP) > 0.95 was taken as the threshold for hypothesis testing.

Epigenome-wide association study (EWAS)

DNA methylation data were regressed onto age, sex, DNAm set, DNAm batch, BMI at Wave 2 (kg/m²), the DNAm-based smoking score EpiSmoker²⁰, four genetic principal components and the measured levels of five immune cells (eosinophils, basophils, lymphocytes, neutrophils and monocytes). EWAS was conducted using OmicS-data-based complex trait analysis (OSCA)³¹. The MOMENT method was used to test for associations between S100β levels and DNAm at individual CpG sites. MOMENT is a mixed-linear-model-based method that is able to account for unobserved confounders and the correlation between distal probes that may be introduced by these

confounders. CpG sites were the independent variables and the dependent variable was the S100 β protein residuals.

Results

Genetic profiling of S100 β

The linear regression genome-wide association study identified 154 SNPs (Figure 2, full summary statistics are available

in the Extended Data) on chromosome 21 that were associated with S100 β levels at $P < 5 \times 10^{-8}$ ($N=769$). There was no evidence of genomic inflation ($\lambda = 0.99$; Figure 3). Conditional and joint analysis (GCTA-COJO) resulted in the identification of one independent pQTL rs8128872 (COJO $\beta = -0.46$, $SE = 0.05$, $P = 3.2 \times 10^{-18}$) associated with S100 β levels. The pQTL was a *cis* variant (located 1,419,889 base

Figure 2. Miami plot of the GWAS (upper panel; $N=769$) and EWAS (lower panel, $N=722$) of S100 β . Blue lines indicate a suggestive threshold of $P < 1 \times 10^{-5}$; red lines indicate genome-wide thresholds of $P < 5 \times 10^{-8}$ (GWAS) and $P < 3.6 \times 10^{-8}$ (EWAS).

Figure 3. QQ Plots and Genomic Inflation statistics for the GWAS (A) and EWAS (B) of S100 β in the LBC1936 sample. Lambda (λ) values are annotated in each case.

pairs downstream of the transcription start site of the *S100β* gene on chromosome 21). There was strong evidence (posterior probability (PP) > 0.95) that two independent causal variants existed for both transcription of *S100β* and *S100β* protein levels in our eQTL analyses for the *S100β* locus (Hypothesis 3: Posterior Probability (PP) = 1.0, Hypotheses 1, 2 and 4 = 0).

Two-sample Mendelian randomisation (MR) was used to test for a causal association between *S100β* serum levels (using our *cis* GWAS data) and Alzheimer's disease. As only one significant SNP remained after LD pruning ($F = 76.67$, indicating a strong effect of the instrument on the *S100β* exposure), the causal effect estimate was determined using the Wald ratio. There was no evidence of an effect of *S100β* serum levels on risk of Alzheimer's disease ($P = 0.95$); (Table 2). Similarly, there was no evidence to suggest that a causal relationship was present between Alzheimer's disease and serum *S100β* ($P > 0.05$ for four of the five methods used). Colocalisation analyses provided further evidence that the loci for Alzheimer's disease and *S100β* were not localised together (Hypothesis 2: Posterior Probability (PP) = 0.99, Hypothesis 3: PP = 0.01, Hypotheses 1 and 4 = 0).

Epigenetic profiling of *S100β*

No CpGs were significantly associated ($P < 3.6 \times 10^{-8}$) with *S100β* levels in the EWAS study ($N=722$) (Figure 2, full summary statistics are available in the Extended Data). The site with the lowest p-value ($P = 5.8 \times 10^{-6}$) was cg06833709, which is located within the *LGII* region, known to encode the leucine-rich glioma inactivated 1 protein (known as epitempin). There was no evidence of genomic inflation ($\lambda = 0.94$, Figure 3).

Discussion

We have characterised the genetic and epigenetic profiles of *S100β*, a circulating protein that has been associated with brain inflammation and neurological disease pathology. We identified a genome-wide significant *cis*-pQTL (rs8128872, $P=5.0 \times 10^{-17}$) that was associated with inter-individual variability in circulating *S100β* levels and found evidence that this pQTL was likely to be distinct from the eQTL for *S100β* transcription. Mendelian randomisation suggested no

evidence of a causal association between *S100β* and Alzheimer's disease or vice versa. Furthermore, there were no CpG probes that had epigenome-wide significant associations with *S100β*.

Whereas the study size was modest for our GWAS and EWAS, previous investigations of similar sample sizes have identified genome-wide SNP and epigenome-wide CpG correlates of protein levels³²⁻³⁵. These include proteomic analyses in the age homogeneous Lothian Birth Cohort 1936 sample that we use in our study^{34,35}. Despite this, larger GWAS and EWAS efforts may help to identify additional loci, which could be used for genetic correlation and more detailed Mendelian randomisation analyses.

Our identification of a single sentinel SNP for *S100β* suggests that though limited, there is some evidence for genetic regulation of this protein in blood. The *cis*-pQTL that we identify (rs8128872) adds to the genetic profile of *S100β* generated in a previous GWAS analysis, which identified two SNPs in the *PCNT* and *HLA* regions¹⁵. While our EWAS suggested that there is no epigenome-wide signature of differential DNAm relating to circulating *S100β* levels, the CpGs with the lowest P values in associations were situated within loci such as *LGII*. The leucine-rich glioma inactivated 1 protein is encoded by *LGII*, primarily locates at neuronal synapses and dysregulation of this protein due to LGII autoimmunity has been directly associated with limbic encephalitis^{36,37}. Whether methylation at this CpG is associated with *S100β* should therefore be confirmed by replication.

The lack of evidence for a causal relationship between *S100β* and Alzheimer's disease suggests either that 1) *S100β* levels in the blood are not directly related to the disease, or 2) any associations are more modest than this study is powered to reliably detect. Studies implicating *S100β* as a candidate marker for dementia are often performed in cerebrospinal fluid^{5,7,8}, which may provide a closer reflection of brain pathology and comparisons between blood and CSF *S100β* levels may therefore yield differing conclusions. However, in the Lothian Birth Cohort 1936 sample we use in this study, serum *S100β* levels have cross-sectionally been associated with poorer general fractional anisotropy⁹, a marker of brain ageing that is associated with increased risk of cognitive decline and dementia³⁸.

Table 2. Mendelian Randomization summary statistics for two-sample, bidirectional tests between *S100β* serum levels and Alzheimer's disease.

Exposure	Outcome	Method	N SNP	Odds Ratio	Beta	SE	P
<i>S100β</i>	Alzheimer's disease	Wald ratio	1	1.0002	3×10^{-4}	4.9×10^{-3}	0.95
Alzheimer's disease	<i>S100β</i>	MR Egger	29	N/A	-0.14	0.51	0.79
Alzheimer's disease	<i>S100β</i>	Weighted median	29	N/A	-0.51	0.51	0.31
Alzheimer's disease	<i>S100β</i>	Inverse variance weighted	29	N/A	-0.77	0.38	0.04
Alzheimer's disease	<i>S100β</i>	Simple mode	29	N/A	-1.07	0.82	0.20
Alzheimer's disease	<i>S100β</i>	Weighted mode	29	N/A	-0.53	0.45	0.24

Given its role as a mediator in inflammatory cascades within the brain^{2,4}, it is likely that S100 β serum levels may be modulated by multiple factors that could be independent of Alzheimer's disease, or indirect from the dementia-associated pathology occurring in the brain. This may be evidenced by our lack of causal association to Alzheimer's disease and these pathways should be explored to delineate targets for therapeutic interventions that may alter neuroinflammation through S100 β mediation.

There are several limitations to our study. First, the Lothian Birth Cohort 1936 are of European ancestries, have little variation in age and selection biases may exist in this cohort, who are considered to be of higher socioeconomic class to the wider Scottish population¹⁶. Therefore, these findings may not generalise to individuals of different ethnic backgrounds, age profiles or socioeconomic groups, though this also means that we are not largely reliant on statistical adjustment for these confounders. Second, our data are from relatively healthy individuals, none of whom reported a diagnosis of dementia at recruitment. It is plausible that analyses in individuals in specific diagnoses groups may yield differing findings. Finally, while the amount of information regarding brain pathology and neurological disease biology is limited when using blood measurements, many blood-based biomarkers have been found to predict and offer insight into Alzheimer's disease³⁹. Therefore, approaches that seek to triangulate between blood, cerebrospinal fluid and brain tissues may strengthen the identification of biomarker signals in future.

Conclusion

We have established evidence for modest genetic, but not epigenetic contributions to the levels of S100 β , a protein marker for brain inflammation and neurological disease. We found no evidence for a causal relationship between serum S100 β and Alzheimer's disease. Future studies should seek to corroborate these findings across blood, cerebrospinal fluid and brain tissue

Data availability

Underlying data

Lothian Birth Cohort 1936 data are not publicly available due to them containing information that could compromise participant consent and confidentiality. Lothian Birth Cohort 1936 data are available on request from the Lothian Birth Cohort Study, University of Edinburgh.

If you are interested in working with the Lothian Birth Cohort 1936 data, you must complete a Data Request Form, indicating

the variables you wish to access from the Data Dictionaries. Data Dictionaries and Data Request Forms are freely accessible at the following website: <https://www.ed.ac.uk/lothian-birth-cohorts/data-access-collaboration>. Completed forms must then be sent to Dr Simon Cox (simon.cox@ed.ac.uk) for approval.

All code is available at the following Gitlab repository: <https://github.com/marioni-group/gwas-and-ewas-of-s100b>.

Extended data

Zenodo: The genetic and epigenetic profile of serum S100 β in the Lothian Birth Cohort 1936 and its relationship to Alzheimer's disease, <https://doi.org/10.5281/zenodo.5591776>.

This project contains the following extended data files:

- s100b_EWAS_output.mlma (EWAS summary statistics for the epigenetic association study of S100 β levels, for each of the 459,309 CpG probes tested)
- s100b_GWAS_output.txt (GWAS summary statistics for the genetic association study of S100 β levels, including the 154 variants that had $P < 5 \times 10^{-8}$)

These are also stored at the following Gitlab repository: <https://github.com/marioni-group/gwas-and-ewas-of-s100b> and will be added to the MRC GWAS and EWAS catalogues.

Reporting guidelines

Zenodo: The STROBE reporting checklist for observational studies, <https://doi.org/10.5281/zenodo.5591776>

Data are available under the terms of the [Creative Commons Zero "No rights reserved" data waiver](#) (CC0 1.0 Public domain dedication).

Ethical approval and consent

Ethical approval for LBC1936 was obtained from the Multi-Centre Research Ethics Committee for Scotland (MREC/01/0/56) and the Lothian Research Ethics committee (LREC/1998/4/183; LREC/2003/2/29). All participants provided written informed consent and the study was performed in accordance with the Helsinki declaration.

Acknowledgements

The authors thank the participants of the Lothian Birth Cohort 1936 study for their valuable contributions to this work.

References

1. Bloomfield SM, McKinney J, Smith L, et al.: **Reliability of S100B in predicting severity of central nervous system injury.** *Neurocrit Care.* 2007; **6**(2): 121–138. [PubMed Abstract](#) | [Publisher Full Text](#)
2. Xia C, Braunstein Z, Toomey AC, et al.: **S100 proteins as an important regulator of macrophage inflammation.** *Front Immunol.* 2018; **8**: 1908. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

3. Venegas C, Heneka MT: **Danger-associated molecular patterns in Alzheimer's disease.** *J Leukoc Biol.* 2017; **101**(1): 87–98.
[PubMed Abstract](#) | [Publisher Full Text](#)
4. Michetti F, Corvino V, Geloso MC, et al.: **The S100B protein in biological fluids: More than a lifelong biomarker of brain distress.** *J Neurochem.* 2012; **120**(5): 644–659.
[PubMed Abstract](#) | [Publisher Full Text](#)
5. Hov KR, Bolstad N, Idland AV, et al.: **Cerebrospinal fluid S100B and Alzheimer's disease biomarkers in hip fracture patients with delirium.** *Dement Geriatr Cogn Dis Extra.* 2017; **7**(3): 374–385.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
6. Chaves ML, Camozzato AL, Ferreira ED, et al.: **Serum levels of S100B and NSE proteins in Alzheimer's disease patients.** *J Neuroinflammation.* 2010; **7**: 6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
7. Peskind ER, Griffin WS, Akama KT, et al.: **Cerebrospinal fluid S100B is elevated in the earlier stages of Alzheimer's disease.** *Neurochem Int.* 2001; **39**(5–6): 409–413.
[PubMed Abstract](#) | [Publisher Full Text](#)
8. Petzold A, Jenkins R, Watt HC, et al.: **Cerebrospinal fluid S100B correlates with brain atrophy in Alzheimer's disease.** *Neurosci Lett.* 2003; **336**(3): 167–170.
[PubMed Abstract](#) | [Publisher Full Text](#)
9. Cox SR, Allerhand M, Ritchie SJ, et al.: **Longitudinal serum S100 β and brain aging in the Lothian Birth Cohort 1936.** *Neurobiol Aging.* 2018; **69**: 274–282.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
10. Zhou S, Bao J, Wang Y, et al.: **S100 β as a biomarker for differential diagnosis of intracerebral hemorrhage and ischemic stroke.** *Neurol Res.* 2016; **38**(4): 327–332.
[PubMed Abstract](#) | [Publisher Full Text](#)
11. Gao Q, Fan Y, Mu LY, et al.: **S100B and ADMA in cerebral small vessel disease and cognitive dysfunction.** *J Neurol Sci.* 2015; **354**(1–2): 27–32.
[PubMed Abstract](#) | [Publisher Full Text](#)
12. Van Eldik LJ, Griffin WS: **S100 beta expression in Alzheimer's disease: relation to neuropathology in brain regions.** *Biochim Biophys Acta.* 1994; **1223**(3): 398–403.
[PubMed Abstract](#) | [Publisher Full Text](#)
13. Gasparoni G, Bultmann S, Lutsik P, et al.: **DNA methylation analysis on purified neurons and glia dissects age and Alzheimer's disease-specific changes in the human cortex.** *Epigenetics Chromatin.* 2018; **11**(1): 41.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
14. Cristóvão JS, Gomes CM: **S100 Proteins in Alzheimer's Disease.** *Front Neurosci.* 2019; **13**: 463.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
15. Wang Y, Nudel R, Benros ME, et al.: **Genome-wide association study identifies 16 genomic regions associated with circulating cytokines at birth.** *PLoS Genet.* 2020; **16**(11): e1009163.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
16. Taylor AM, Pattie A, Deary IJ: **Cohort profile update: The Lothian birth cohorts of 1921 and 1936.** *Int J Epidemiol.* 2018; **47**(4): 1042r.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
17. Deary IJ, Gow AJ, Taylor MD, et al.: **The Lothian Birth Cohort 1936: a study to examine influences on cognitive ageing from age 11 to age 70 and beyond.** *BMC Geriatr.* 2007; **7**: 28.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
18. Davies G, Tenesa A, Payton A, et al.: **Genome-wide association studies establish that human intelligence is highly heritable and polygenic.** *Mol Psychiatry.* 2011; **16**(10): 996–1005.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
19. Shah S, McRae AF, Marioni RE, et al.: **Genetic and environmental exposures constrain epigenetic drift over the human life course.** *Genome Res.* 2014; **24**(11): 1725–1733.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
20. Bollepalli S, Korhonen T, Kaprio J, et al.: **EpiSmoker: A robust classifier to determine smoking status from DNA methylation data.** *Epigenomics.* 2019; **11**(13): 1469–1486.
[PubMed Abstract](#) | [Publisher Full Text](#)
21. McCartney DL, Stevenson AJ, Hillary RF, et al.: **Epigenetic signatures of starting and stopping smoking.** *EBioMedicine.* 2018; **37**: 214–220.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
22. Trejo Banos D, McCartney DL, Patxot M, et al.: **Bayesian reassessment of the epigenetic architecture of complex traits.** *Nat Commun.* 2020; **11**(1): 2865.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
23. R. C. T.: **R: A language and environment for statistical computing.** R Foundation for Statistical Computing, Vienna, Austria, 2017.
[Reference Source](#)
24. Purcell S, Neale B, Todd-Brown K, et al.: **PLINK: A tool set for whole-genome association and population-based linkage analyses.** *Am J Hum Genet.* 2007; **81**(3): 559–575.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
25. Yang J, Ferreira T, Morris AP, et al.: **Conditional and joint multiple-SNP analysis of GWAS summary statistics identifies additional variants influencing complex traits.** *Nat Genet.* 2012; **44**(4): 369–375, S1–3.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
26. Vösa U, Claringbould A, Westra HJ, et al.: **Large-scale cis- and trans-eQTL analyses identify thousands of genetic loci and polygenic scores that regulate blood gene expression.** *Nat Genet.* 2021; **53**(9): 1300–1310.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
27. Guo H, Fortune MD, Burren OS, et al.: **Integration of disease association and eQTL data using a Bayesian colocalisation approach highlights six candidate causal genes in immune-mediated diseases.** *Hum Mol Genet.* 2015; **24**(12): 3305–3313.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
28. Giambartolomei C, Vukcevic D, Schadt EE, et al.: **Bayesian Test for Colocalisation between Pairs of Genetic Association Studies Using Summary Statistics.** *PLoS Genet.* 2014; **10**(5): e1004383.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
29. Jansen IE, Savage JE, Watanabe K, et al.: **Genome-wide meta-analysis identifies new loci and functional pathways influencing Alzheimer's disease risk.** *Nat Genet.* 2019; **51**(3): 404–413.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
30. Hemani G, Zheng J, Elsworth B, et al.: **The MR-base platform supports systematic causal inference across the human phenome.** *eLife.* 2018; **7**: e34408.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
31. Zhang F, Chen W, Zhu Z, et al.: **OSCA: A tool for omic-data-based complex trait analysis.** *Genome Biol.* 2019; **20**(1): 107.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
32. Sun BB, Maranville JC, Peters JE, et al.: **Genomic atlas of the human plasma proteome.** *Nature.* 2018; **558**(7708): 73–79.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
33. Zaghlool SB, Kühnel B, Elhadad MA, et al.: **Epigenetics meets proteomics in an epigenome-wide association study with circulating blood plasma protein traits.** *Nat Commun.* 2020; **11**(1): 15.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
34. Hillary RF, Trejo-Banos D, Kousathanas A, et al.: **Multi-method genome- And epigenome-wide studies of inflammatory protein levels in healthy older adults.** *Genome Med.* 2020; **12**(1): 60.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
35. Hillary RF, McCartney DL, Harris SE, et al.: **Genome and epigenome wide studies of neurological protein biomarkers in the Lothian Birth Cohort 1936.** *Nat Commun.* 2019; **10**(1): 3160.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
36. Zekeridou A, Pittock SJ: **Synaptic autoimmunity: new insights into LGI1 antibody-mediated neuronal dysfunction.** *Brain.* 2020; **143**(6): 1622–1625.
[PubMed Abstract](#) | [Publisher Full Text](#)
37. Wegner F, Wilke F, Raab P, et al.: **Anti-leucine rich glioma inactivated 1 protein and anti-N-methyl-D-aspartate receptor encephalitis show distinct patterns of brain glucose metabolism in ¹⁸F-fluoro-2-deoxy-d-glucose positron emission tomography.** *BMC Neurol.* 2014; **14**: 136.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
38. Power MC, Su D, Wu A, et al.: **Association of white matter microstructural integrity with cognition and dementia.** *Neurobiol Aging.* 2019; **83**: 63–72.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
39. Lantero Rodriguez J, Karikari TK, Suárez-Calvet M, et al.: **Plasma p-tau181 accurately predicts Alzheimer's disease pathology at least 8 years prior to post-mortem and improves the clinical characterisation of cognitive decline.** *Acta Neuropathol.* 2020; **140**(3): 267–278.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Open Peer Review

Current Peer Review Status:

Version 1

Reviewer Report 17 December 2021

<https://doi.org/10.21956/wellcomeopenres.19151.r47163>

© 2021 Sharma M. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Manu Sharma

Centre for Genetic Epidemiology, Institute for Clinical Epidemiology and Applied Biometry, University of Tübingen, Tübingen, Germany

The identification of relevant biomarker signatures for age-related neurodegenerative diseases is an area that merits well-designed studies. The study by Gadd *et al.* is such an effort in which the authors have performed genome and epigenome-wide association studies (GWAS/EWAS) of serum S100 β levels in older adults. Subsequently, they also performed Mendelian randomization (MR) with Alzheimer's disease (AD) to infer a causal relationship between S100 β levels and AD.

It is a well-designed study. The methodology applied and subsequent interpretation of their findings are well interpreted. I do not have any major comments regarding their study design and interpretation.

Minor comments:

The authors should explain in detail their MR design and analysis. The study cohort is comparatively small. The authors should explain this as a caveat in the discussion. It could probably well be their effect estimates and subsequent p-values are suffering from the "winner's curse". The independent cohort validation is a must before any meaningful inferences can be deduced from the present study.

Is the work clearly and accurately presented and does it cite the current literature?

Yes

Is the study design appropriate and is the work technically sound?

Yes

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Neurogenetics, Genetic Epidemiology, Statistical Genetics.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 27 Jan 2022

Danni Gadd, University of Edinburgh, Edinburgh, UK

Thank you for raising these important points on the MR analysis.

We have added further clarification regarding the MR analyses. The new text that has been added to the methods section is summarised here for reference:

'Associations from separate GWAS were used as genetic instruments. As allele assignment is randomised, the SNPs associated with the exposure are randomised to the effects of confounders and likely to be causally upstream of the exposure...

...The GWAS summary statistics for S100 β were sourced from the analyses in this study that used samples from the Lothian Birth Cohort 1936 (N=769). Importantly, the Alzheimer's disease summary statistics were based on a meta-analysis of cohorts that were independent of the Lothian Birth Cohort 1936...

...One assessment quantified the association between S100 β levels as the exposure and Alzheimer's disease as the outcome. A second assessment then quantified the association between Alzheimer's disease as the exposure and S100 β levels and the outcome...

...The effect estimate for S100 β to Alzheimer's disease was therefore determined using the Wald ratio test (a ratio of effect per risk allele on trait to effect per risk allele on protein levels)...

...As multiple independent variants were identified, a multi-method MR approach was chosen. Unity between the estimates from these methods indicates that the results are more likely to be robust. The MR Egger approach did not find strong evidence of horizontal pleiotropy present (non-significant MR-Egger intercept).'

Upon revising the manuscript, we also noticed a discrepancy in how the effect allele frequencies were loaded into the Mendelian randomisation analysis pipeline for our

Alzheimer's disease to S100 β levels direction of MR analysis and have rectified this. The updated results are included in the revised results table, which differ marginally from those included previously and still indicate no significant associations from Alzheimer's disease to S100 β levels.

We also agree that the GWAS dataset is relatively small and have added a caveat in our discussion to emphasise that independent validation is required to confirm the results we have generated in MR in future work. This key update reads as follows:

'The GWAS sample size was relatively small and it is imperative that our GWAS and Mendelian randomisation results are independently validated by future cohorts that have S100 β measurements available. This will further elucidate the likelihood of a causal relationship between S100 β and Alzheimer's disease.'

We also state the following in the discussion:

'...larger GWAS and EWAS efforts may help to identify additional loci, which could be used for genetic correlation and more detailed Mendelian randomisation analyses.'

We hope that by making our summary statistics and code open access, there will be further cohorts with S100 β measurements that can assess the replicability of these findings.

Competing Interests: N/A

Reviewer Report 14 December 2021

<https://doi.org/10.21956/wellcomeopenres.19151.r47417>

© 2021 Wood N. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Nicholas W. Wood

Department of Clinical and Movement Neurosciences, UCL Queen Square Institute of Neurology, University College London, London, UK

An interesting and important study to evaluate the possible biologically motivated causal relationship between S100 and Alzheimer's disease. Too often biomarkers are reported for a variety of neurodegenerative diseases and no definite (or often plausible) biological link is made. Here, the authors have used 2-stage Mendelian randomisation to attempt to investigate whether SNPs associated with S100 levels and/or a methylation epigenetic readout are causally related to Alzheimer's disease.

They first performed an analysis in the Lothian birth cohort to 'build' the eugenic instruments and then applied them to the very large GWAS datasets available for Alzheimer's. They do not find any evidence that genetically encoded determination of S100 and its methylation is linked to AD. This

is useful to the community.

My minor comments:

- Could they detail what amount of the variance of expression is captured by the instrument?
- Also, I think for clarity, explaining a bit more about the trio probability assessments in the Coloc analyses would help the non-specialist reader.

Is the work clearly and accurately presented and does it cite the current literature?

Yes

Is the study design appropriate and is the work technically sound?

Yes

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

Partly

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Neurogenetics

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 27 Jan 2022

Danni Gadd, University of Edinburgh, Edinburgh, UK

Thank you for identifying these opportunities to add further clarification to the results.

We have added the variance in S100 β levels that could be explained by the lead variant (rs8128872, $r^2 = 9\%$) in our revision that has been submitted.

We have also clarified the individual hypotheses in the colocalisation analyses and added further explanation regarding the rationale of the probability assessment when performing colocalisation between traits. New text that has been added to this section is provided below for reference:

'Colocalisation analysis can be used to derive the probability that common genetic variants are shared between two phenotypes in a given region of the genome...

...Rare variants with $MAF < 0.01$ and variants with missing MAF were excluded from the analysis. A total of 1,346 variants were included for Alzheimer's disease and 1,010 variants were included for $S100\beta$. A single causal variant assumption is made in the analysis that there is one causal variant per trait and the probability of colocalisation between loci can be derived...

...In addition to the null hypothesis (no causal variants in the region assessed), hypothesis 1 indicated a causal variant was present for Alzheimer's disease only. Hypothesis 2 indicated that there was a causal variant for $S100\beta$ levels only. Hypothesis 3 indicated that there were independent causal variants for both Alzheimer's disease and $S100\beta$ levels. Hypothesis 4 indicated that a common variant contributed to both Alzheimer's disease and $S100\beta$ levels.'

Competing Interests: N/A